

# **Contribution of Inherited Variants to Hereditary Cancer Syndrome Predisposition**

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Cancer is a clonal disease that develops as a result of the changes on the genetic material by various factors in micro/macro environment. It has a multi-step development process. In some cancer types, genetic factors allow this multi-step process to proceed easily. These cancer types are also called hereditary cancer syndromes. Targeted gene panels are important diagnostic methods in hereditary cancer syndromes to detect the causative variants associated with these hereditary cancer syndromes. We reviewed the data of 94 patients who applied to Ankara City Hospital Genetic Diseases Evaluation Center from March 2019 to July 2021. Qiagen familial cancer susceptibility gene panel kit was used for next generation sequencing to detect the single nucleotide variants for the targeted genes. Sixty-one genes which are associated with increased cancer risk or well characterized hereditary cancer syndromes were included to this panel. Twenty five patients (27%), including 8 males and 17 females, had pathogenic/likely pathogenic variants in 13 of the 61 genes analyzed. Forty patients (43%) had variants which were assessed as variant of unknown significant. In our study, targeted multi-gene panel was diagnostic in nearly one third of the patients with personal/familial cancer syndromes. Molecular diagnosis in familial cancer syndromes is important in terms of predictive diagnosis and family screening, as well as patient follow-up and early prophylactic surgery. The predisposition for hereditary cancer syndromes can be determined according to pre-test evaluation, figuring out the inheritance type with pedigree analysis, cancer type and the genetic analysis for appropriate susceptibility genes.

**Keywords:** genetics; hereditary cancer syndrome; next generation sequencing; pathogenic variant; variant of unknown significance

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### Introduction

Cancer is a clonal disease that develops as a result of the changes on the genetic material by various factors in micro/macro environment with a multi-step development process. In some cancer types, genetic factors allow this multi-step process to begin easily. In this case, it is noted that there is a "genetic predisposition" for cancer. These cancer types are also referred to as hereditary cancer syndromes. Hereditary cancers make up 5-10% of all cancer types. Most of them are inherited as autosomal dominant (American College of Obstetricians and Gynecologists 2019). Up to now, more than 200 hereditary cancer syndromes have been reported (Machačkova et al. 2016; Tsaousis et al. 2019).

Targeted gene panels are important diagnostic methods in hereditary cancer syndromes (LaDuca et al. 2014). There are many advantages of targeted next generation sequencing (NGS) gene panels utilization in hereditary cancer syndromes, including cost-effectiveness or time saving (Pritchard et al. 2012; LaDuca et al. 2014). Morbidity and mortality can be reduced by the correct genetic diagnosis in patients with hereditary cancer syndromes and the preventive procedures can be performed to the patients. The family members that are at risk can also be tested, and genetic counseling can be given to them (according to the test results) (de Oliveira et al. 2022).

This study aims to detect the frequency of germline

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pathogenic/likely pathogenic variants in cancer susceptibility genes by the utilization of targeted NGS panels for the patients or family members referred with hereditary cancer syndromes.

#### **Materials and Methods**

#### Patients

We reviewed the data of 94 patients as two groups who applied to Ankara City Hospital Genetic Diseases Evaluation Center from March 2019 to July 2021. One group of patients had a history of different solid tumors affecting more than one system in themselves and/or their families, while the other group underwent BRCA1/2 deletion/duplication analysis for breast-ovarian cancer and had no pathogenicity. The patients were referred to our genetic laboratory for diagnostic genetic test.

Permission for the study was obtained from Ankara City Hospital Ethics Committee (E1-22-2488). The study followed the guidelines and principles of the Declaration of Helsinki. All patients and formal guardians of the patients under 18 signed the written informed consent for the usage of their clinical data and genetic analysis.

Genomic DNA was extracted from peripheral blood using QIAcube® automatic DNA isolation system (QIAGEN Inc., Mississauga, ON, Canada) according to the manufacturer's instructions. Qiagen familial cancer susceptibility gene panel CDHS-13974Z-2393 kit (QIAGEN, Hilden, Germany) was used for NGS to detect the single nucleotide variants for the targeted genes. The target enrichment process was followed by sequencing of the libraries on Illumina MiSeq sysytem (Illumina Inc., San Diego, CA, USA).

#### Data analysis and variant interpretation

Data analysis was carried out by QIAGEN Clinical Insight (QCITM) software (QIAGEN). Pathogenic, likely pathogenic, and uncertain significant variants were confirmed by Sanger sequencing. The exons of all targeted genes were sequenced at a read depth of  $30 \times$  or greater. 2015 American College of Medical Genetics Standards and Guidelines (ACMG) were used for the interpretation of sequence variants 10 (Richards et al. 2015).

#### Results

A total of 94 patients (33 males and 61 females) with suspected hereditary cancer syndromes were analysed for molecular etiology. The range of ages of the patients was between 2-71. Thirty three male cases had a mean age of 45.7; and 61 female cases had a mean age of 40.2. Sixteen patients were in the pediatric age group. Sixty-one genes which are associated with increased cancer risk or well characterized hereditary cancer syndromes were included to this panel (Table 1).

Out of 94 cases referred to us for genetic testing, 90 patients (96%) had a personal cancer history. The majority of the patients had a personal history of colon adenocarcinoma (14%) and breast cancer (14%). A family history for cancer was positive for 49% (46/94) of the whole patients. Pathogenic variants were detected in 21 of these 46 patients with a positive family history, and variant of unknown significant (VUS) were detected in 19 of them, six patients had no pathogenic variant/VUS. Missense variants were the most prevalent variation type, accounting for the 40% (10/25) of the variants identified, followed by nonsense, frameshift, insertion/deletion variations and splicing variations.

Twenty five patients (27%), including 8 males and 17 females, had pathogenic/likely pathogenic variants in 13 of the 61 genes analyzed (TP53, CHEK2, MSH2, MUTYH, MMLH1, SDHB, VHL, BLM, BRCA2, MRE11, MSH6, PTCH1 and RAD51D genes) (Fig. 1a). Five patients (19%) were under 18 years old. Family history was positive at twenty two patients out of twenty five patients who had pathogenic/likely pathogenic variants. Only two patients had no positive family history, and both of them had pathogenic variant at SDHB gene that has autosomal dominant inheritance. While one of these patients had paraganglioma, the other had pheochromocytoma. The other pathogenic variants were at TP53 (4), CHEK2 (3), MSH2 (3), MUTYH (3), MMLH1 (2), VHL (2), BLM (1), BRCA2 (1), MRE11 (1), MSH6 (1), PTCH1 (1) and RAD51D (1) genes (Fig. 1b). One patient diagnosed with colon cancer had compound heterozygosity for MUTYH gene. Another patient diagnosed with lymphoblastic lymphoma had homozygote pathogenic variant at MSH2 gene. The family histories, ages, and sex of the patients, genes and pathogenic/ likely pathogenic variants are shown in Table 2.

BRCA1/2 MLPA (Multiplex Ligation-dependent Probe Amplification) were only performed on seven patients with breast cancer, three patients with ovarian cancer and one patient with pancreas cancer because of the high frequency of BRCA deletion/duplications in these cancer types. No variants were found. With targeted gene NGS, two patients had pathogenic variants at TP53 and RAD51D genes, four patients had VUS at APC, PALB2, RAD50 and PTCH1 genes. Five patients had no variations.

Forty patients (43%) had variants which were assessed as variant of unknown significant in our study (Fig. 1a). VUS was detected at NBN (1), PMS1 (2), BUB1B (2), CTNNA1 (1), SDHD (2), ATR (4), STK11 (1), RAD50 (3), POLE (3), RET (1), GEN1 (1), PTCH1 (2), FLCN (1), ATM (2), RAD51D (1), MSH6 (1), POLD1 (2), NTHL1 (1), PALB2 (2), SMARCA4 (2), CDH1 (2), MRE11 (2), BRIP1 (1), PMS2 (1), BARD1 (1), RAD51C (1) and APC (2) genes. Two patients had two and three VUS in different genes. These patients were followed up and called for annual controls. Pathogenic, likely pathogenic and VUS were confirmed by bidirectional Sanger sequencing. Over 99% of the coding exons of all genes in the panel were sequenced to a read depth of  $30 \times$  or greater in almost all cases. According to these results, molecular diagnosis rate was 27%.

Gene Name	Transcript ID	Inheritance	Gene Name	Transcript ID	Inheritance
AIP	NM_003977.4	AD	MSH2	NM_000251.3	AD, AR
APC	NM_000038.6	AD	MSH6	NM_000179.3	AD, AR
ATM	NM_000051.4	AD, AR	MUTYH	NM_001048174.2	AR
ATR	NM_001184.4	AD	NBN	NM_002485.5	AR
AXIN2	NM_004655.4	AD	NTHL1	NM_002528.7	AR
BAP1	NM_004656.4	AD	PALB2	NM_024675.3	AD
BARD1	NM_000465.4	AD	PALLD	NM_001166108.2	AD
BLM	NM_000057.4	AR	PIK3CA	NM_006218.4	-
BMPR1A	NM_004329.3	AD	PMS1	NM_000534.5	-
BRCA1	NM_007294.4	AD	PMS2	NM_000535.5	AR
BRCA2	NM_000059.4	AD, AR	POLD1	NM_002691.3	AD
BRIP1	NM_032043.3	AD	POLE	NM_006231.3	AD
BUB1B	NM_001211.6	-	PRSS1	NM_002769.5	AD
CDH1	NM_004360.5	AD	PTCH1	NM_000264.5	AD
CDK4	NM_000075.4	AD	PTEN	NM_000314.8	AD
CDKN2A	NM_058195.4	AD	RAD50	NM_005732.4	AR
CHEK2	NM_007194.4	AD	RAD51B	NM_133510.4	-
CTNNA1	NM_001903.5	AD	RAD51C	NM_058216.2	AR
EPCAM	NM_002354.3	AD	RAD51D	NM_002878.3	-
FAM175A	-	-	RET	NM_020975.4	AD
FANCC	NM_000136.3	AR	RINT1	NM_021930.6	AR
FLCN	NM_144997.7	AD	SDHB	NM_003000.2	AD
GALNT12	NM_024642.5	-	SDHC	NM_003001.3	AD
GEN1	NM_001130009.3	-	SDHD	NM_003002.3	AD
GPC3	NM_004484.4	XLR	SMAD4	NM_005359.5	AD
GREM1	NM_013372.7	-	SMARCA4	NM_003072.5	AD
HOXB13	NM_006361.6	-	STK11	NM_000455.4	AD
MEN1	NM_001370259.2	AD	<i>TP53</i>	NM_000546.5	AD
MET	NM_000245.4	AD	VHL	NM_000551.3	AD
MLH1	NM_000249.4	AD, AR	XRCC2	NM_005431.2	AR
MRE11A	NM_005591.4	AR			

Table 1. Hereditary cancer syndrome-associated genes (n = 61) included in targeted next generation sequencing (NGS) panel, their corresponding transcript numbers and heredity types.

AD, autosomal dominant; AR, autosomal recessive; XLR, X-linked-recessive.

#### Discussion

Multi-gene panels using NGS technology make it possible to identify pathogenic variants at hereditary cancer susceptibility genes. Each mutated gene included in the panel can increase the risk for hereditary cancer syndomes (de Oliveira et al. 2022). Thus, these multi-gene panels ensure the risk estimation in predictive medicine (Colas et al. 2019). In this study, hereditary cancer related pathogenic variants were detected by using NGS technology at the susceptibility genes for hereditary cancers. A total of 94 individuals who were referred to our clinics for hereditary cancer syndromes were included in our study. All of the individuals had personal/family history. The genetic analysis disclosed the presence of at least one pathogenic/likely pathogenic variant in 27% of the individuals, and VUS were detected in 43% of the cases. Thus, the contribution of all genes in our panel was important for the hereditary cancer predisposition. The diagnosis rate was similar to previous studies (Tsaousis et al. 2019; Ercoskun et al. 2022).

Pathogenic variants were detected mostly at patients with colon adenocarcinoma (24%), following ovarian cancer (12%), brain tumors (12%), breast cancer (8%), paraganglioma (8%) and less at patients with other types of cancer. Among the patients with a pathogenic variant, 16% (4/25) of the cases had variants in the *TP53* gene, indicating a significant contribution of this gene in cancer predisposition. Apart from the *TP53* gene, the other highly mutated genes were *CHEK2* (12%), *MUTYH* (8% monoallelic variants, 4% biallelic variants), and *MSH2* (12%). At least one pathogenic/likely pathogenic variant was identified in 25 of



Fig. 1. Distribution of variants found in the patient group (a), and the genes in which the pathogenic variants were detected (b).VUS, variant of unknown significant.

samples (27%) including all of the individuals with no information about their personal/family cancer history.

The rate of hereditary cancer syndromes in adults is between 5-10%, but in children it has not been fully investigated. In a study by Knapke et al. (2012) at a pediatric cancer clinic, the patients were followed for over 2-years. They were screened for family history, age, ethnicity and tumor characteristics underlying cancer predisposition syndrome. Pathogenic variants were detected at 29% of the children due to the family history with cancer (Knapke et al. 2012; Schiffman et al. 2013). In our study, the diagnosis rate in pediatric age group was 37.5%, while it was 24% in adulthood period, compatible with the literature.

Biallelic pathogenic variants of MUTYH gene are associated with a syndrome known as MUTYH associated polyposis (MAP) syndrome (Tsaousis et al. 2019). In only one case (a 66 years old female patient with colon adenocarcinoma), MUTYH variations were detected as compound heterozygote in our study. Monoallelic MUTYH pathogenic variant is also significant as a risk factor for colorectal cancer. The risk is 2.5 times more according to general population (Win et al. 2014). Two cases in our study were heterozygote for different monoallelic variations of MUTYH gene. One of the patients was diagnosed with breast cancer and the other had recurrent endometrial polyp with positive family histories, respectively. MUTYH heterozygotes may have a mildly elevated risk for colorectal carcinoma according to NCCN guidelines (NCCN, The National Comprehensive Cancer Network 2018). There was no other multiple pathogenic variant found in any of our patients except from MUTYH.

Pathogenic variants of *TP53* are associated with a wide range of cancers, collectively known as Li Fraumeni Syndrome, which is characterised by a predisposition to some malignancies like adrenocortical carcinomas, soft-tissue sarcomas, early-onset breast cancer, brain tumors and

leukaemias (Paduano et al. 2021, 2022). *TP53* had the highest rate (16%) for pathogenic variant in our study. The patients had different cancer types (adrenocortical carcinoma, brain tumor, breast and colon cancer) with positive family history, in line with the literature.

*SDHB* pathogenic variants were detected in two patients with paraganglioma in our study. Succinate dehydrogenase genes (*SDHA*, *SDHB*, *SDHC*, *SDHD*, and *SDHAF2*) are associated with hereditary paraganglioma syndromes in literature. For paraganglioma syndromes, to identify a pathogenic variant is important for the further surveillance protocol recommendations to the patients and their families. This actually illustrates the importance of analyzing susceptibility gene variants in familial cancer syndromes and the importance of the diversity of the genes included in the panel for different familial cancer types (Horton et al. 2022).

A pathogenic missense variant in *BLM* gene was detected at 44 years old female patient who was diagnosed with colon adenocarcinoma. Her sister had renal cell carcinoma, her cousin had lung adenocarcinoma and her grandmother had pancreas cancer. de Voer et al. (2015) reported two patients carried a pathogenic *BLM* variant in a cohort of 55 early-onset colorectal carcimoma cases ( $\leq$  45 years old). Pathogenic *BLM* variations may cause an increased risk for developing colorectal carcinoma at an early age, but some other risk factors are also important for the carcinogenesis period (de Voer et al. 2015).

The overall rate of VUS in our study was around 43-45% similar to other studies with a maximum of two VUS per individual (Tsaousis et al. 2019; Ercoskun et al. 2022). The highest VUS rate among individuals with personal history of cancer was detected with breast cancer (12.5%), colon adenocarcinoma (10.0%) and pancreas cancer (10.0%). Only one unaffected individual with family history of ovarian cancer had VUS at *CDH1* gene. Family

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Table 2. Twenty-five cases with pathogenic/likely pathogenic variants and family histories.

F, Female; M, Male; HGMD, Human Gene Mutation Database.

study may also be recommended at VUS variants, but it may not provide complete information because of the incomplete penetrance and variable expression of cancer susceptibility (Garrett et al. 2016; Anderson et al. 2022). Patients with VUS were called for annual controls. Since the age of onset is different in patients, it is appropriate to evaluate the patients with VUS once a year (Colas et al. 2019). This is because accurate VUS assessment needs a professional review of literature using ACMG guidelines (Anderson et al. 2022). As the data gets bigger, specific revisions for disease or the variants are required (Nykamp et al. 2017; Waddell-Smith et al. 2020) In addition to this, as the number of genes included in the panel increases, there will be a higher VUS risk (de Oliveira et al. 2022).

Molecular diagnosis in familial cancer syndromes is important in terms of predictive diagnosis and family screening, as well as patient follow-up and early prophylactic surgery. Selecting panels containing target genes will enable patients to receive molecular diagnosis in a shorter time. Studying large multigene panels will contribute to the diagnosis in cases with atypical phenotypic features or when the etiology cannot be clarified with single gene-targeted studies. It is also recommended that laboratories performing multiple gene testing share their datas, thereby contributing to the increase of this useful information.

In our study, targeted multi-gene panel was diagnostic in nearly one third of the patients with personal/familial cancer syndromes. The predisposition for hereditary cancer syndromes can be determined according to pre-test evaluation, deciding the inheritance type with pedigree analysis, cancer type and the genetic analysis for appropriate susceptibility genes. Variable onset of the disease, personal cancer or positive family history are important for a correct genetic diagnosis. An accurate diagnosis is also important for an appropriate genetic counseling for the patients to understand the significance of genetic testing.

## **Conflict of Interest**

The authors declare no conflict of interest.

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