



# CircRNA CircABC1 Diminishes the Sensitivity of Breast Cancer Cells to Docetaxel by Sponging MiR-153-3p

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Resistance to docetaxel is a major problem to the success of docetaxel-based therapies for breast cancer. The present study was to identify the role of circABC1 in altering the docetaxel resistance properties. Reverse transcription-quantitative PCR (qRT-PCR) was performed to quantify circABC1 and miR-153-3p. The effects of circABC1 on the viability, apoptosis and migration/invasion of docetaxel-resistant and -sensitive cells were investigated by cell function experiments, including Cell Counting Kit-8 and Transwell assays. Correlation between circABC1 and the docetaxel-treated outcome was analyzed by multivariate Cox regression analysis, in addition to Kaplan-Meier analysis of time to treatment failure (TTF). The targeting relationship between circABC1 and miR-153-3p was predicted and verified by dual-luciferase reporter assay and RNA immunoprecipitation. CircABC1 was highly expressed in cancerous tissues, as well as the docetaxel-sensitive group and cells. The overexpression of circABC1 contributed to cell viability, docetaxel-resistance and migration/invasion, but inhibited apoptosis. CircABC1 can sponge miR-153-3p. CircABC1 contributed to the docetaxel resistance of breast cancer, maybe via the miR-153-3p.

**Keywords:** breast cancer; ceRNA; circABC1; docetaxel resistance; prognosis

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## Introduction

In 2020, there were 19.29 million new cancer cases in the world, of which 2.26 million were breast cancer. Breast cancer has surpassed lung cancer (2.2 million) to become the largest cancer in the world for the first time (Siegel et al. 2022). Among the 4.43 million female cancer deaths, 680,000 were caused by breast cancer, ranking first (Wilkinson and Gathani 2022). In China, according to the data of the China urban cancer registry, the overall cancer incidence rate has increased by 20%-30% in the past 30 years and has increased by 3%-5% every year (Lei et al. 2021). This growth rate is significantly higher (1.5%) than the average world growth rate. The choice of treatment for breast cancer is usually based on lymph node metastasis and stage (Maughan et al. 2010). Choice for early-stage (I and II) breast cancers is usually breast-conserving surgery and radiation therapy (Curigliano et al. 2017). Chemotherapy and endocrine therapy are usually consid-

ered for node-positive breast cancer. Induction chemotherapy is used to treat stage III breast cancer to down-size the tumor. Treatment options for women with recurrent or metastatic (stage IV) should consider the benefits of life length and the harm of pain (Akram et al. 2017). Notably, drug resistance to targeted and systemic therapies is one of the causes of breast cancer-related mortality (Rivera and Gomez 2010). The development of chemoresistance remains a major bottleneck in the treatment of cancer patients.

In the past two decades, our understanding of the role of noncoding RNA (ncRNA) in gene regulation and cell biology has been steadily improving. ncRNA contains various types of RNAs, such as microRNAs (miRNAs) and circular RNAs (circRNAs). The discovery of circRNAs has brought our understanding of the important role of these RNA molecules to a new stage. First, lncRNAs can play a role in many biological processes, such as cell proliferation/differentiation/growth, development, metabolism, aging,

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and cell death (Li et al. 2020; Ghafouri-Fard et al. 2021). Secondly, circRNAs can act as key regulators in cancer, such as oncogenes or tumor suppressors, which may affect the development, metastasis and chemoresistance of many different types of cancers, including breast cancer (Lei et al. 2020; Li et al. 2020). In addition, lncRNAs can act as bait to competitively bind to miRNAs and thus affect the expression and function of downstream genes, which are defined as competing endogenous RNAs (ceRNAs) (Tay et al. 2014; Li et al. 2020). Some circRNAs are related to the prevalence of drug resistance phenotype and the recovery of drug sensitivity of cancer cells (Li et al. 2020). For instance, dysregulation of circKDM4C and hsa\_circ\_0006528 can mitigate or contribute, respectively, to doxorubicin resistance in breast cancer (Yu et al. 2015; Liang et al. 2019). At present, docetaxel is a common Taxus drug, which plays an important role in the chemotherapy of breast cancer. CircCYP24A1, circDPP4, and circ-XIAP have been proven to promote the development of docetaxel resistance in cancers (Zhang et al. 2021; Gu and Duan 2022; Yin et al. 2022). Therefore, targeting circRNAs may become a new strategy for the treatment of drug-resistant cancers.

CircABC1 has been screened as a dysregulated circRNA in docetaxel-resistant breast cancer cell sublines (Huang et al. 2021). In this study, we further investigated the expression levels of circABC1 in normal tissues and docetaxel-resistant/sensitive breast cancer tissues. Subsequently, the biological function, function mechanism, and clinical value of circABC1 in breast cancer were explored based on clinical specimens, pathological information, and *in vitro* experiments.

## Materials and Methods

### Patients

The patients in the Zibo Central Hospital between March 2019 and March 2020 with locally advanced or metastatic breast cancer received docetaxel as a first-line chemotherapy drug, not combined with other drugs. The patient who had relapsed cancer or received neoadjuvant chemotherapy before inclusion were excluded. A total of 90 women were included in the study who met the Response Evaluation Criteria in Solid Tumors (Eisenhauer et al. 2009). The surgical material from patients was collected upon written informed consent. Docetaxel-resistant (DOCR) tissue samples (n = 40) and docetaxel-sensitive (DOCS) tissue samples (n = 50) were acquired from patients. The procedures used in this study adhere to the tenets of the Declaration of Helsinki. The present study was approved by the Ethics Committee of Zibo Central Hospital, and informed consent was obtained from all subjects involved in the study. Time to treatment failure (TTF) was defined as the period from the beginning of treatment to disease progression, loss of follow-up or replacement, or interruption of treatment for any reason.

### Cell culture

Docetaxel-resistant human breast cancer cell line, MDA-MB-231/DOCR, was purchased from Ybio (Shanghai, China). Its parental cell line, MDA-MB-231, along with BT-474, SK-BR-3, and MCF7, was obtained from National Biomedical Experimental Cell Resource Bank (Beijing, China). These cell lines were cultured in DMEM (Gibco, Palo Alto, CA, USA) supplemented with 10% fetal calf serum (FCS) (Gibco). MDA-MB-231/DOCR cells were maintained in a medium with 65 nM docetaxel. The cultures were kept in a humidified atmosphere with 5% carbon dioxide at 37°C.

### Cell transfection

An appropriate amount of cells were inoculated into cell culture dishes. After the cells adhered to the wall, the cells were washed, and the culture medium without FCS was added. The pcDNA, siRNA, and miRNA mimics used for transfection were designed and synthesized by RIBO Biology. Lipofectamine 3000 (Invitrogen, Waltham, MA, USA) was used for transfection. After 8 h of transfection, the medium containing FCS was added to replace the previous medium. After 24 h of transfection, the follow-up cell function test was started; At the same time, a small number of cells were taken to extract RNA, for verification of the transfection efficiency by qRT-PCR.

### Total RNA purification and quantification of circABC1 and miR-153-3p

The cell culture dish was taken out, and then the culture medium was discarded. The patient's tumor tissues and their adjacent normal tissues were transferred to eppendorf tube, and ground. Total RNA of cells and tissues was isolated and purified using the RNAiso Plus (TAKARA, Kyoto, Japan). When circABC1 was reverse transcribed, PrimeScript RT Master Mix kit was used; When miR-153-3p was reverse transcribed, the Mir-X miRNA First Strand Synthesis kit was used. The target molecular expression levels were detected using SYBR green Primeix ExTaq II kit according to the product manual. GAPDH served as an endogenous control for circABC1, while U6 for miR-153-3p. The relative expression levels were calculated with the  $2^{-\Delta\Delta Ct}$  approach.

### Detection of IC50

The IC50 values of docetaxel for MDA-MB-231 and MDA-MB-231/DOCR cells were determined. Using 96-well plates (Sigma-Aldrich, Sigma-Aldrich, MO, USA), MDA-MB-231 cells were seeded at  $5 \times 10^4$  cells per well; MDA-MB-231/DOCR, at  $3 \times 10^4$  cells per well. After 24 h, cells were exposed to relevant docetaxel solutions of different concentrations (0, 1, 2, 4, 6, 8, 10, and 15  $\mu\text{g}/\text{mL}$ ) for 24 h. CCK-8 kit was used to determine the corresponding optical density values according to the instructions from the manufacturer. IC50 values were calculated by non-linear regression.

### Cell migration and invasion

The transfected MDA-MB-231 and MDA-MB-231/DOCR cells were counted, washed twice, and resuspended in FCS-free medium at concentrations of  $9 \times 10^4/200 \mu\text{L}$  and  $5 \times 10^4/200 \mu\text{L}$ , respectively. On the 24-well cell plate, a complete medium containing 20% FCS (700  $\mu\text{L}$  per hole) was added. The transwell inserts with (for cell invasion) or without (for cell migration) Matrigel were put into the 24 well plates and filled with the prepared cell suspensions. The culture time of MDA-MB-231 was 36 h, and that of MDA-MB-231/DOCR was 20 h. After culture, the cells were washed twice, fixed with methanol, and dyed with 0.1% crystal violet. After removing the cells in the upper chamber, the migrated cells were counted then.

### Cell apoptosis analysis

According to the product manual, Annexin V-FITC Propidium Iodide (PI) apoptosis detection kit was used to analyze apoptosis. MDA-MB-231 and MDA-MB-231/DOCR cells were transfected for 24 h. Cell suspension was prepared using Annexin V Binding Solution. 5  $\mu\text{L}$  Annexin V/FITC solution/5  $\mu\text{L}$  PI solution was made to incubate the cells darkly at room temperature for 15 min. FACS Calibur was used for the experiment, and Flowjo 7.6.1 analyzed the data of apoptosis experiment.

### Bioinformatics analysis

The targeting miRNAs for circABC1 were predicted by Circular RNA Interactome (<https://circinteractome.nia.nih.gov/index.html>) database. The predicted miRNA with the highest 'context+ score percentile' was selected for verification.

### Dual-luciferase reporter assay

For verification of the targeting relationship, dual-luciferase reporter assay was used. The vectors were used to construct wild-type reporter plasmid of circABC1 (wt-circ) and mutant plasmid of circABC1 (site 1 mut, site 2 mut, and site 1 + 2 mut). Next, MDA-MB-231/DOCR cells were co-transfected with miR-153-3p mimic or its negative control (NC) and the reporter plasmids using Lipofectamine 3000 reagent (Invitrogen). After 48 h, the relative luciferase activity was determined by a Dual Luciferase Reporter Gene Assay Kit.

### RNA binding protein immunoprecipitation (RIP) assay

Magna RIP™ RNA-Binding Protein Immunoprecipitation Kit (Merck Millipore, Burlington, MA, USA) was obtained to determine the connectivity between circABC1 and miR-153-3p. Briefly, the cell pellet was re-suspended in an equal pellet volume of complete RIP Lysis Buffer. The Argonaute2 monoclonal antibody (Ago2) was immunoprecipitated to RNA-binding protein (RBP) of interest with protein A/G magnetic beads. The beads-bounding complexes were immobilized with a magnetic magnet and the unbound materials were washed off. RNAs were extracted

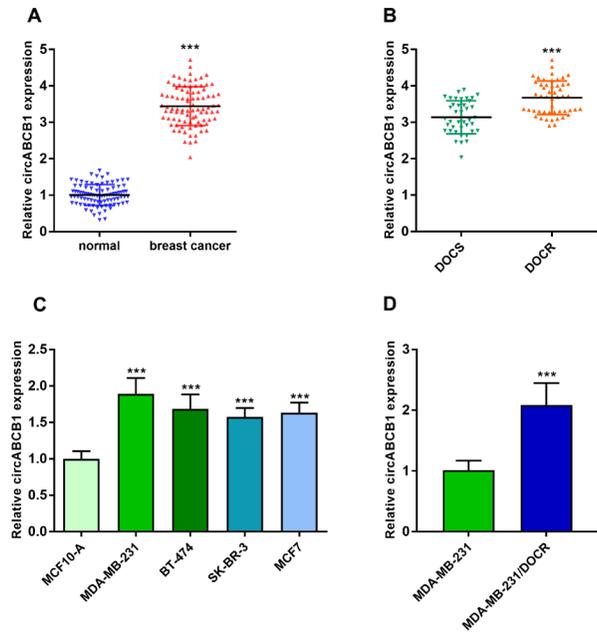


Fig. 1. Expression of circABC1 in tissues and cells.

(A, B) The expression of circABC1 was quantified in normal breast tissues and cancerous tissues, including docetaxel-resistant and -sensitive groups. (C, D) The expression of circABC1 was quantified in normal mammary epithelial cells and cancerous cells, including docetaxel-resistant and -sensitive groups. DOCR, docetaxel-resistant; DOCS, docetaxel-sensitive. \*\*\* $P < 0.001$ .

and subjected to qRT-PCR.

### Statistics

All analyzed data were from triplicate or more experiments. According to whether the data conformed to the Gaussian distribution, the data between the two groups were compared by unpaired t-test or Mann Whitney U test. For multiple-group comparisons, analysis of variance (ANOVA) followed by Dunnett method was employed. To test the association between two quantitative variables, the Pearson correlation was used. The nonrandom association of co-occurrence between two categorical variables was tested by the Fisher's exact test. Multivariate Cox regression and Kaplan-Meier survival analyses were utilized to identify prognostic significance of circABC1 for time to treatment failure (TTF).  $P$  value  $< 0.05$  was the boundary of significance or not.

## Results

### Docetaxel-resistant tissues and cells showed increased expression of circABC1 compared to docetaxel-sensitive ones

To evaluate the expression of circABC1, we used qRT-PCR in normal and cancerous tissues. We found that circABC1 was highly expressed in cancerous tissues compared to normal tissues (Fig. 1A). Further, we got an

Table 1. Association of the circABC1 expression with clinical characteristics in patients with advanced breast cancer.

Variables	Cases (n = 90)	CircABC1 ≤ median	CircABC1 > median	P
Age				
≤ 48	46	24	22	0.673
> 48	44	21	23	
HER2				
negative	65	29	36	0.099
positive	25	16	9	
HR				
negative	69	35	34	0.803
positive	21	10	11	
Metastasis sites				
none	52	32	20	0.028
viscera	23	9	14	
others	15	4	11	
TNM stage				
III	50	29	21	0.090
IV	40	16	24	
Histological grade				
I	56	25	31	0.247
II	26	14	12	
III	8	6	2	

HER2, human epidermal growth factor receptor 2; HR, hormone receptor (estrogen receptor or progesterone receptor).

Table 2. Multivariable analysis of time to treatment failure.

Variables	HR (95%CI)	P
CircABC1 (> median vs. ≤ median)	2.892 (1.352-6.189)	0.006
Age (≤ 48 years vs. > 48 years)	1.442 (0.760-2.738)	0.263
HER2 (negative vs. positive)	1.213 (0.576-2.551)	0.612
HR (negative vs. positive)	1.115 (0.518-2.400)	0.780
Metastasis sites		
Viscera vs. none	2.159 (0.933-4.995)	0.072
Others vs. none	1.895 (0.727-4.936)	0.191
TNM stage (IV vs. III)	1.610 (0.758-3.420)	0.216
Histological grade		
II vs. I	2.657 (1.296-5.448)	0.008
III vs. I	4.074 (1.515-10.956)	0.005

HER2, human epidermal growth factor receptor 2; HR, hormone receptor (estrogen receptor or progesterone receptor).

upregulation of the expression level of circABC1 in docetaxel-resistant tissues when compared to the docetaxel-sensitive ones (Fig. 1B). In breast cancer cell lines, circABC1 also exhibited a higher expression than the normal cells (Fig. 1C). Given that MDA-MB-231 presented the highest level of circABC1, we chose MDA-MB-231 and

its docetaxel-resistant cell line (MDA-MB-231/DOCR) for subsequent experiments. As expected, circABC1 expression exhibited significant differences between the docetaxel-sensitive (lower) and -resistant (higher) cells (Fig. 1D). These results indicate that circABC1 was upregulated in breast cancer, especially docetaxel-resistant breast cancer.

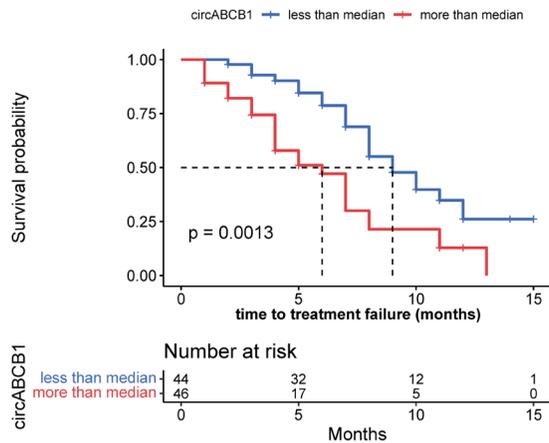


Fig. 2. CircABC1 was related to a shorter time to treatment failure (TTF).

The figure shows circABC1 expression split into two expression categories using the median expression value and its relationship with TTF in a Kaplan-Meier graph (log-rank test).

### CircABC1 levels were associated with TTF

We next assessed whether circABC1 expression was associated with the outcome of patients who received docetaxel therapy. To this end, we evaluated the contribution of circABC1 using multivariate Cox regression analysis and Kaplan-Meier survival analysis. In the Pearson correlation analysis, the high expression of circABC1 (> median value) was associated with metastasis (Table 1). In the Cox regression analysis, we evaluated the contribution of circABC1 in a multivariate model, and circABC1 was associated with the periods of TTF (HR = 2.892, 95% CI 1.352-6.189,  $P = 0.006$ ) (Table 2). The association of circABC1 with TTF was visualized by a Kaplan-Meier plot after dividing our cohort into circABC1-high and -low subgroups based on the median expression level (Fig. 2). These results indicated that circABC1 was an independent predictive marker for poor TTF.

### CircABC1 regulated the viability of docetaxel-resistant and -sensitive MDA-MB-231 cells

To identify circRNAs related to docetaxel sensitivity or resistance, MDA-MB-231 cells were transfected with

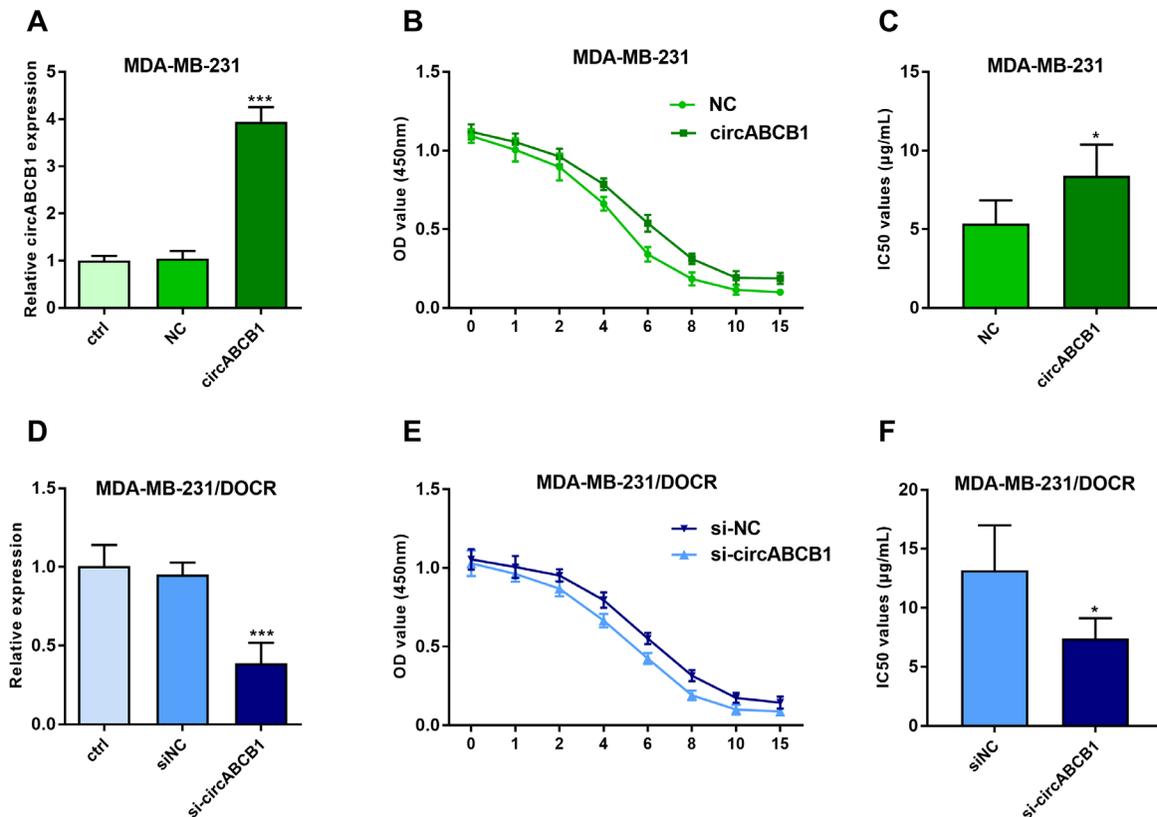


Fig. 3. CircABC1 influenced the viability of docetaxel-resistant and -sensitive cells.

(A) qRT-PCR analysis of circABC1 levels in transfected MDA-MB-231 cells. (B) CCK-8 viability assay was carried out in the circABC1-transfected MDA-MB-231 cells. (C) IC50 was determined for MDA-MB-231 cells. (D) qRT-PCR analysis of circABC1 levels in transfected MDA-MB-231/DOCR cells. (E) CCK-8 viability assay was carried out in the circABC1-knockdown MDA-MB-231/DOCR cells. (F) IC50 was determined for MDA-MB-231/DOCR cells. DOCR, docetaxel-resistant. \* $P < 0.05$ , \*\*\* $P < 0.001$ .

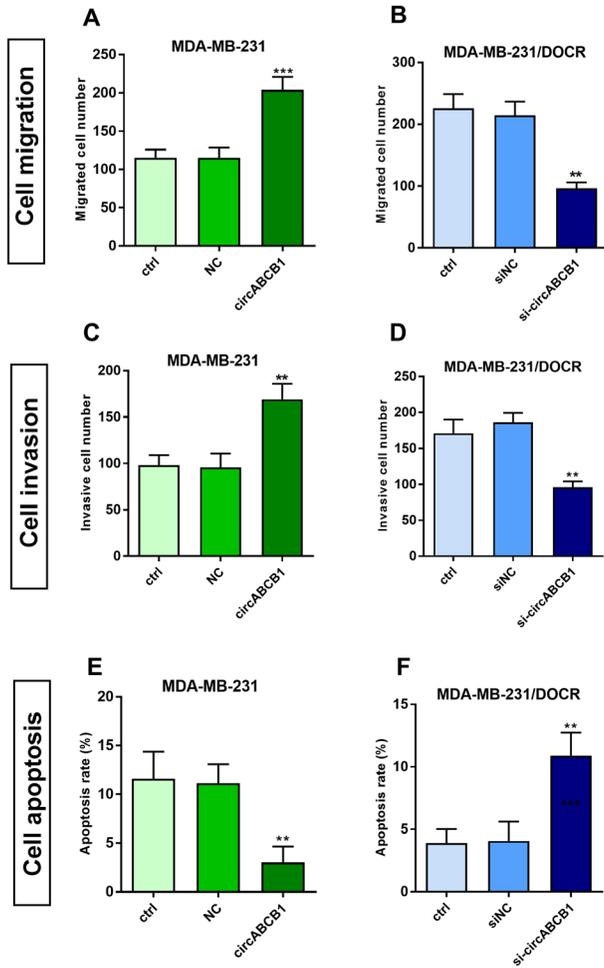


Fig. 4. CircABC B1 affects the apoptotic and migratory capacities of docetaxel-resistant and -sensitive MDA-MB-231 cells. (A, B) The cell migration assay. (C, D) The cell invasion assay. (E, F) The cell apoptosis assay. DOCR, docetaxel-resistant. \*\* $P < 0.01$ , \*\*\* $P < 0.001$ .

circABC B1 overexpression plasmid, and MDA-MB-231/DOCR cells were transfected with circABC B1 siRNA (si-circABC B1). qRT-PCR results confirmed the transfection efficiency in MDA-MB-231 cells (Fig. 3A), determining an increase of circABC B1 in cells transfected by circABC B1 overexpression plasmid. The CCK-8 assay indicated that overexpressed circABC B1 enhanced the viability of parental MDA-MB-231 cells (Fig. 3B) and increased the IC<sub>50</sub> value (Fig. 3C). The transfection of MDA-MB-231/DOCR was also successful (Fig. 3D). The silencing of circABC B1 significantly reduced the viability of MDA-MB-231/DOCR cells (Fig. 3E) and decreased the IC<sub>50</sub> value (Fig. 3F).

#### *CircABC B1 affected the apoptotic and migratory capacities of docetaxel-resistant and -sensitive MDA-MB-231 cells*

It was demonstrated in Transwell migration assay that

the overexpression of circABC B1 distinctly increased the number of migratory MDA-MB-231 cells (Fig. 4A), while knockdown of circABC B1 reduced the number of migratory MDA-MB-231/DOCR cells (Fig. 4B). Similarly, an increased number of invasive MDA-MB-231 cells was observed after the overexpression of circABC B1 (Fig. 4C), whereas a suppression after the silencing of circABC B1 (Fig. 4D). The apoptotic assay demonstrated that the overexpression of circABC B1 decreased the percentage of apoptotic parental MDA-MB-231 cells (Fig. 4E), whereas the silence of circABC B1 promoted apoptosis of the docetaxel-resistant MDA-MB-231/DOCR cells (Fig. 4F).

#### *Prediction of the interactions of circABC B1 with miR-153-3p*

Given the fact that circRNA can bind miRNA through their miRNA response elements (MREs), we predicted the targeting miRNA of circABC B1. CircABC B1 contained two MREs for miR-153-3p (Fig. 5A). MiR-153-3p was listed first in candidate target miRNAs with 'context+ score percentile' of 96 and 86. MiR-153-3p was lowly expressed between the normal tissues and cancerous tissues (Fig. 5B), as well as the docetaxel-sensitive and -resistant cell groups (Fig. 5C). So was the expression in cell lines (Fig. 5D, E). The expression levels of circABC B1 and miR-153-3p appeared to be a negative association (Fig. 5F). Subsequently, the results of the dual luciferase reporter assay (Fig. 5G) and RIP assay (Fig. 5H) verified that circABC B1 directly targeted miR-153-3p.

## Discussion

Taxanes (including docetaxel and paclitaxel) have become the first-line treatment drugs for breast cancer (Sachdev and Jahanzeb 2016). However, genetic or acquired drug resistance hinders their effectiveness, and there are few biomarkers for predicting Taxanes resistance (Murray et al. 2012). As a chemotherapy drug for patients with breast cancer, docetaxel is widely used in clinical practice; however, more and more patients produce docetaxel resistance (Sekino and Teishima 2020). Many studies have explored the mechanism of the docetaxel-resistance. Among them, the study on the mechanism of epigenetic modification occupies a place in drug resistance-related mechanisms (Jia et al. 2019; Garcia-Martinez et al. 2021). Epigenetic modification is a heritable genetic change that affects DNA expression without changing gene nucleotide sequence. It includes DNA methylation, histone modification, and regulation of various ncRNAs (Miranda Furtado et al. 2019). CircRNA is recognized as a new class of ncRNA that plays an important role in the occurrence and progression of breast cancer (Tang et al. 2021). But its role in docetaxel resistance of breast cancer patients is less studied. In this study, we first measured circABC B1 in docetaxel-resistant breast cancer tissues. The results showed that the expression of circABC B1 in docetaxel-resistant tissues was significantly higher than that in

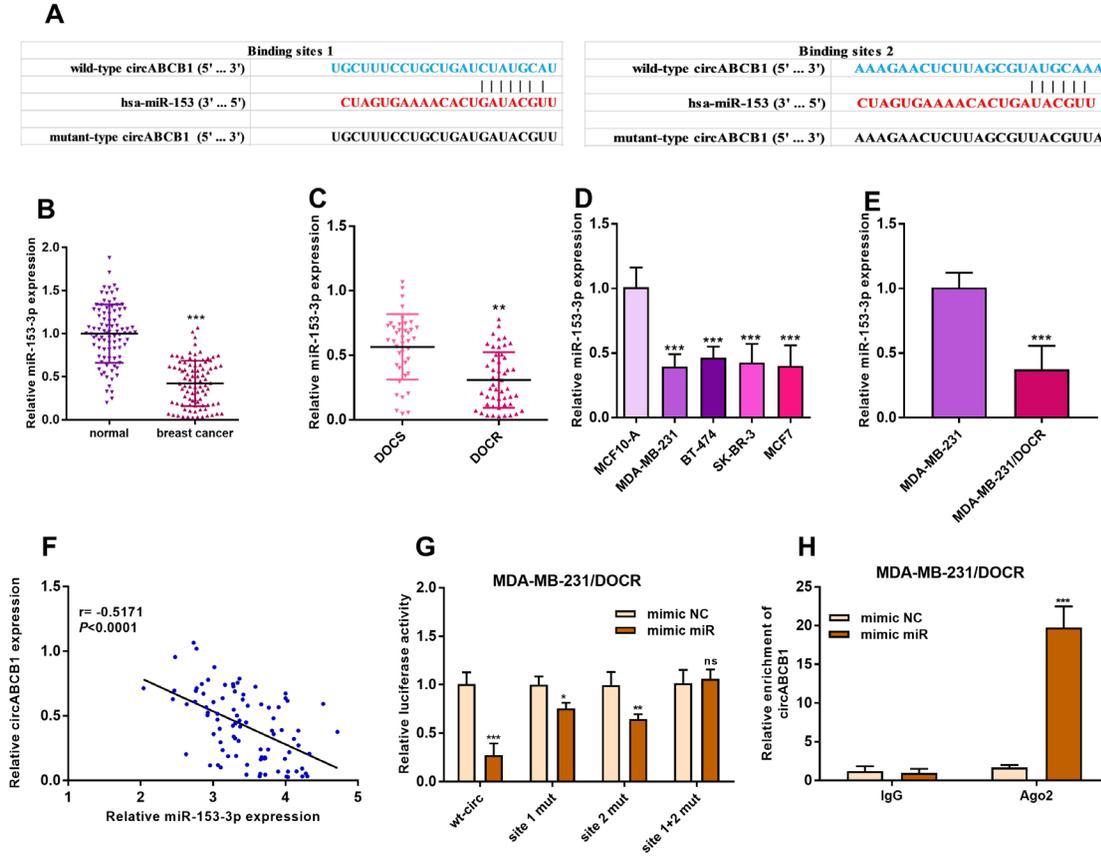


Fig. 5. CircABC1 interacted with miR-153-3p.

(A) Target prediction analysis of circABC1 and miR-153-3p. (B-E) The relative expression level of miR-153-3p in tissues and cells. (F) The correlation between circABC1 expression and miR-153-3p expression in cancerous tissues. (G) Luciferase reporter assay. (H) RNA Binding Protein Immunoprecipitation (RIP) Assay. Ago2, Argonaute2 monoclonal antibody; DOCR, docetaxel-resistant; DOCS, docetaxel-sensitive; ns, not significant. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ .

docetaxel-sensitive tissues. The expression level of MDA-MB-231/DOCR in drug-resistant cell line was also significantly higher than that in parent cell line. We also found that the expression level of circABC1 in tissue samples was positively correlated with the tumor metastasis of patients through the correlation analysis between the expression of circABC1 in samples and clinical pathological parameters of patients. Based on the follow-up information, we found that patients with high circABC1 expression showed shorter TTF. Through overexpression and interference experiments, we found that circABC1 has the biological functions of increasing IC<sub>50</sub> of docetaxel, promoting tumor cell migration/invasion, and inhibiting apoptosis. These results suggest that circABC1 not only participates in the regulation of chemotherapy resistance of docetaxel in breast cancer patients, but also may play an important role in the occurrence and development of tumors as a tumor-promoting factor.

It has been reported that the most common mechanism of circRNA in tumor cells is to act as a “sponge” for miRNA, regulating the expression and activity of its down-

stream target genes by adsorbing miRNA (Han et al. 2018). CiRS-7 is the first reported circRNA with miRNA “sponge” function, which has more than 70 miR-7 binding sites (Hansen et al. 2013; Weng et al. 2017). Subsequent studies found that miR-7 was adsorbed through the “sponge” mechanism of circRNA molecules (Weng et al. 2017). CiRS-7 could reduce the activity of miR-7 and improve the expression of a series of downstream target genes, such as *EGFR*, *RAF1*, *E2A*, *NF-κB*, and *HOXB13*, and then perform biological functions. In addition to ciRS-7, many other circRNAs also play an important role in different malignant tumors through the above mechanism. CircRNA-100290 promotes the occurrence and development of oral squamous cell carcinoma by adsorbing miR-378a (Chen et al. 2019); circHIPK3 sensitizes paclitaxel-resistant breast cancer cells to paclitaxel therapy at least partly via the regulation of the miR-1286 (Ni et al. 2021); CircLARP4 has been proved to inhibit the proliferation and invasion of tumor cells by adsorbing miR-424-5p in gastric cancer (Zhang et al. 2017). In this subject, we found that circABC1 can bind to miR-153-3p and affect its expression through bioinformatics

prediction, RIP and double luciferase reporter gene detection experiments. Previous studies have confirmed that miR-153-3p plays an anti-tumor role in breast cancer and inhibits the proliferation and migration of breast cancer cells (Sun et al. 2020). It is worth noting that overexpression of miR-153-3p can inhibit the drug resistance of triple-negative breast cancer cells to paclitaxel by inducing G2/M phase (Wang et al. 2020). This suggests that miR-153-3p may be a potential therapeutic target for paclitaxel-resistant patients. Therefore, the above results suggest that miR-153-3p, as an important functional target gene of circABC1, may mediate the promotion of circABC1 in breast cancer and docetaxel resistance in breast cancer.

To sum up, this study shows that circABC1 plays an important role in promoting breast cancer and participates in the regulation of docetaxel drug sensitivity. *In vitro* experiments confirmed that circABC1 overexpression can significantly inhibit cell proliferation/migration, and increase the proportion of apoptosis and sensitivity to tamoxifen. Therefore, the above experimental results suggest that circABC1 may be a new therapeutic target for breast cancer patients, especially those who use docetaxel as a first-line drug. In addition, its regulatory effect on docetaxel resistance suggests that circABC1 can also be used as a new molecular marker to predict docetaxel resistance in breast cancer patients, which has important clinical significance. However, as for the application of circABC1 in clinical practice, further patient follow-up data should be collected and analyzed with a larger sample size.

### Conflict of Interest

The authors declare no conflict of interest.

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### References

- Akram, M., Iqbal, M., Daniyal, M. & Khan, A.U. (2017) Awareness and current knowledge of breast cancer. *Biol. Res.*, **50**, 33.
- Chen, X., Yu, J., Tian, H., Shan, Z., Liu, W., Pan, Z. & Ren, J. (2019) Circle RNA hsa\_circRNA\_100290 serves as a ceRNA for miR-378a to regulate oral squamous cell carcinoma cells growth via Glucose transporter-1 (GLUT1) and glycolysis. *J. Cell. Physiol.*, **234**, 19130-19140.
- Curigliano, G., Burstein, H.J., Winer, E.P., Gnant, M., Dubsky, P., Loibl, S., Colleoni, M., Regan, M.M., Piccart-Gebhart, M., Senn, H.J., Thurlimann, B.; St. Gallen International Expert Consensus on the Primary Therapy of Early Breast Cancer 2017, Andre, F., Baselga, J., Bergh, J., et al. (2017) De-escalating and escalating treatments for early-stage breast cancer: the St. Gallen International Expert Consensus Conference on the Primary Therapy of Early Breast Cancer 2017. *Ann. Oncol.*, **28**, 1700-1712.
- Eisenhauer, E.A., Therasse, P., Bogaerts, J., Schwartz, L.H., Sargent, D., Ford, R., Dancey, J., Arbuck, S., Gwyther, S., Mooney, M., Rubinstein, L., Shankar, L., Dodd, L., Kaplan, R., Lacombe, D., et al. (2009) New response evaluation criteria in solid tumours: revised RECIST guideline (version 1.1). *Eur. J. Cancer*, **45**, 228-247.
- Garcia-Martinez, L., Zhang, Y., Nakata, Y., Chan, H.L. & Morey, L. (2021) Epigenetic mechanisms in breast cancer therapy and resistance. *Nat. Commun.*, **12**, 1786.
- Ghafari-Fard, S., Shoorei, H., Bahroudi, Z., Abak, A. & Taheri, M. (2021) The role of H19 lncRNA in conferring chemoresistance in cancer cells. *Biomed. Pharmacother.*, **138**, 111447.
- Gu, H. & Duan, Z. (2022) Silencing of circDPP4 suppresses cell progression of human prostate cancer and enhances docetaxel cytotoxicity through regulating the miR-564/ZIC2 axis. *J. Gene Med.*, **24**, e3403.
- Han, B., Chao, J. & Yao, H. (2018) Circular RNA and its mechanisms in disease: from the bench to the clinic. *Pharmacol. Ther.*, **187**, 31-44.
- Hansen, T.B., Kjems, J. & Damgaard, C.K. (2013) Circular RNA and miR-7 in cancer. *Cancer Res.*, **73**, 5609-5612.
- Huang, P., Li, F., Mo, Z., Geng, C., Wen, F., Zhang, C., Guo, J., Wu, S., Li, L., Brunner, N. & Stenvang, J. (2021) A comprehensive RNA study to identify circRNA and miRNA biomarkers for docetaxel resistance in breast cancer. *Front. Oncol.*, **11**, 669270.
- Jia, Z.H., Wang, X.G. & Zhang, H. (2019) Overcome cancer drug resistance by targeting epigenetic modifications of centrosome. *Cancer Drug Resist.*, **2**, 210-224.
- Lei, M., Zheng, G., Ning, Q., Zheng, J. & Dong, D. (2020) Translation and functional roles of circular RNAs in human cancer. *Mol. Cancer*, **19**, 30.
- Lei, S., Zheng, R., Zhang, S., Wang, S., Chen, R., Sun, K., Zeng, H., Zhou, J. & Wei, W. (2021) Global patterns of breast cancer incidence and mortality: a population-based cancer registry data analysis from 2000 to 2020. *Cancer Commun. (Lond.)*, **41**, 1183-1194.
- Li, J., Sun, D., Pu, W., Wang, J. & Peng, Y. (2020) Circular RNAs in cancer: biogenesis, function, and clinical significance. *Trends Cancer*, **6**, 319-336.
- Liang, Y., Song, X., Li, Y., Su, P., Han, D., Ma, T., Guo, R., Chen, B., Zhao, W., Sang, Y., Zhang, N., Li, X., Zhang, H., Liu, Y., Duan, Y., et al. (2019) circKDM4C suppresses tumor progression and attenuates doxorubicin resistance by regulating miR-548p/PBLD axis in breast cancer. *Oncogene*, **38**, 6850-6866.
- Maughan, K.L., Lutterbie, M.A. & Ham, P.S. (2010) Treatment of breast cancer. *Am. Fam. Physician*, **81**, 1339-1346.
- Miranda Furtado, C.L., Dos Santos Luciano, M.C., Silva Santos, R.D., Furtado, G.P., Moraes, M.O. & Pessoa, C. (2019) Epidrugs: targeting epigenetic marks in cancer treatment. *Epigenetics*, **14**, 1164-1176.
- Murray, S., Briasoulis, E., Linardou, H., Bafaloukos, D. & Papadimitriou, C. (2012) Taxane resistance in breast cancer: mechanisms, predictive biomarkers and circumvention strategies. *Cancer Treat. Rev.*, **38**, 890-903.
- Ni, J., Xi, X., Xiao, S. & Xiao, X. (2021) Silencing of circHIPK3 sensitizes paclitaxel-resistant breast cancer cells to chemotherapy by regulating HK2 through targeting miR-1286. *Cancer Manag. Res.*, **13**, 5573-5585.
- Rivera, E. & Gomez, H. (2010) Metastasis resistance in metastatic breast cancer: the evolving role of ixabepilone. *Breast Cancer Res.*, **12** Suppl 2, S2.
- Sachdev, J.C. & Jahanzeb, M. (2016) Use of cytotoxic chemotherapy in metastatic breast cancer: putting taxanes in perspective. *Clin. Breast Cancer*, **16**, 73-81.
- Sekino, Y. & Teishima, J. (2020) Molecular mechanisms of docetaxel resistance in prostate cancer. *Cancer Drug Resist.*, **3**, 676-685.
- Siegel, R.L., Miller, K.D., Fuchs, H.E. & Jemal, A. (2022) Cancer statistics, 2022. *CA Cancer J. Clin.*, **72**, 7-33.
- Sun, L., Wang, H., Jiang, J. & Bi, X. (2020) miR-153-3p inhibits proliferation and migration of breast cancer cells via down-regulating ROCK1. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi*, **36**, 138-144 (in Chinese).

- Tang, L., Jiang, B., Zhu, H., Gao, T., Zhou, Y., Gong, F., He, R., Xie, L. & Li, Y. (2021) The biogenesis and functions of circRNAs and their roles in breast cancer. *Front. Oncol.*, **11**, 605988.
- Tay, Y., Rinn, J. & Pandolfi, P.P. (2014) The multilayered complexity of ceRNA crosstalk and competition. *Nature*, **505**, 344-352.
- Wang, Y., Wu, N., Zhang, J., Wang, H. & Men, X. (2020) MiR-153-5p enhances the sensitivity of triple-negative breast cancer cells to paclitaxel by inducing G2M phase arrest. *Oncotargets Ther.*, **13**, 4089-4097.
- Weng, W., Wei, Q., Toden, S., Yoshida, K., Nagasaka, T., Fujiwara, T., Cai, S., Qin, H., Ma, Y. & Goel, A. (2017) Circular RNA ciRS-7-A promising prognostic biomarker and a potential therapeutic target in colorectal cancer. *Clin. Cancer Res.*, **23**, 3918-3928.
- Wilkinson, L. & Gathani, T. (2022) Understanding breast cancer as a global health concern. *Br. J. Radiol.*, **95**, 20211033.
- Yin, H., Qin, H., Yang, L., Chen, M., Yang, Y., Zhang, W., Hao, J., Lu, Q., Shi, J., Zhuang, J., Qiu, X. & Guo, H. (2022) circCYP24A1 promotes docetaxel resistance in prostate cancer by upregulating ALDH1A3. *Biomark. Res.*, **10**, 48.
- Yu, D.D., Wu, Y., Shen, H.Y., Lv, M.M., Chen, W.X., Zhang, X.H., Zhong, S.L., Tang, J.H. & Zhao, J.H. (2015) Exosomes in development, metastasis and drug resistance of breast cancer. *Cancer Sci.*, **106**, 959-964.
- Zhang, H., Li, M., Zhang, J., Shen, Y. & Gui, Q. (2021) Exosomal circ-XIAP promotes docetaxel resistance in prostate cancer by regulating miR-1182/TPD52 axis. *Drug Des. Devel. Ther.*, **15**, 1835-1849.
- Zhang, J., Liu, H., Hou, L., Wang, G., Zhang, R., Huang, Y., Chen, X. & Zhu, J. (2017) Circular RNA\_LARP4 inhibits cell proliferation and invasion of gastric cancer by sponging miR-424-5p and regulating LATS1 expression. *Mol. Cancer*, **16**, 151.
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