

Immunohistochemical Demonstration of Somatostatin-Containing Cells in the Human, Dog and Rat Thyroids

YUKIO YAMADA, SEIKI ITO, YOHICHI MATSUBARA* and SHIGERU KOBAYASHI†

*The First Department of Internal Medicine, *The First Department of Surgery and †The Third Department of Anatomy, Niigata University School of Medicine, Niigata*

YAMADA, Y., ITO, S., MATSUBARA, Y. and KOBAYASHI, S. *Immunohistochemical Demonstration of Somatostatin-Containing Cells in the Human, Dog and Rat Thyroids.* Tohoku J. exp. Med., 1977, 122 (1), 87-92 — By an indirect immunofluorescence technique using the somatostatin antibody, somatostatin-containing cells were demonstrated to exist in the human, dog and rat thyroids. These cells were located predominantly in the interfollicular areas and more scatteringly in the follicular areas. The source of these somatostatin-containing cells was discussed. ——— somatostatin; thyroid; immunofluorescence technique; parafollicular cells

Somatostatin (growth hormone release-inhibiting hormone) was originally isolated from the ovine hypothalamus (Brazeau et al. 1973), later characterized (Burgus et al. 1973) and then synthesized (Rivier et al. 1973). Both natural and synthetic somatostatin preparations inhibit not only the release of growth hormone (GH) (Brazeau et al. 1973; Lovinger et al. 1973; Siler et al. 1973; Vale et al. 1972, 1974) but also the release of thyrotropin (TSH) (Hall et al. 1973; Siler et al. 1973; Vale et al. 1974) from the pituitary gland. Furthermore, somatostatin also suppresses the release of extrapituitary hormones such as insulin, glucagon (Alberti et al. 1973; Alford et al. 1974; Koerker et al. 1974; Mortimer et al. 1974; Sakurai et al. 1974; Yen et al. 1974) and gastrin (Bloom et al. 1974).

In addition to the studies on the physiological effects of somatostatin mentioned above, Arimura et al. (1975a) prepared antibodies to this substance and this advance has allowed immunohistochemical and radioimmunoassay methods to be employed for the search of localization of somatostatin in tissues; it is demonstrable not only in the central nervous system (Hökfelt et al. 1974, 1975; Pelletier et al. 1974; Dubois et al. 1975a; King et al. 1975; Brownstein et al. 1975; Alpert et al. 1976), but also in the endocrine pancreas and the gastro-intestinal tract (Luft et al. 1974; Dubois 1975; Dubois et al. 1975b; Polak et al. 1975; Arimura et al. 1975b). Furthermore, Hökfelt et al. (1975) have recently demonstrated with fluorescent histochemistry the occurrence of somatostatin-containing cells in the rat thyroid.

Although this report suggested the existence of somatostatin-containing cells in the thyroid of many mammals including human, the proof remained to be obtained.

In this study we demonstrate, by the indirect immunofluorescence technique, the occurrence of somatostatin-containing cells in the thyroid of the human, dog and rat.

MATERIALS AND METHODS

Antibody preparation

Somatostatin antibody. Synthetic somatostatin (Peptide Institute Research Foundation, Osaka, Japan) used as the antigen was coupled with bovine serum albumin (BSA) by the use of glutaraldehyde. The somatostatin-BSA complex was emulsified with complete Freund's adjuvant and injected into rabbits subcutaneously at intervals of 30–40 days.

Immunofluorescent agents. Flouresecein isothionate (FITC)-conjugated anti-rabbit IgG goat serum (Miles, England) was used.

Tissue preparation

Fresh thyroids were removed from the human, dog and rat, and small blocks were fixed in Bouin's solution at room temperature for 3 hr. Then they were dehydrated and embedded in paraffin wax (melting point 42–44°C). Sections were cut (5 μ m thick) and fixed on glass slides treated with a mixture of 0.5% gelatin and 0.05% chromium sulfate.

Immunofluorescence

After removal of paraffin wax with xylene, a drop of somatostatin antibody diluted 1:20 with a phosphate-buffered saline solution (PBS, pH 7.4, 0.05 M) was applied to each section, which was then incubated for 2 hr at 37°C in a moist chamber. All the sections were then washed in the buffer described above, and anti-rabbit IgG serum-FITC diluted with PBS was applied to them before they were again incubated for 1 hr at 37°C in a moist chamber.

Control study

As controls for somatostatin antibody specificity, either 1:20 dilution of normal rabbit serum or 1:20 dilution of somatostatin antibody absorbed with 500 μ g of synthetic somatostatin, 100 μ g of thyroxine, 100 μ g of triiodothyronine or 100 μ g of porcine calcitonin per ml of undiluted antibody was substituted for the specific antibody in the staining procedure.

RESULTS

Somatostatin-containing cells were found in the human (Fig. 1), dog (Figs. 2, 3) and rat (Fig. 4) thyroids, although the number of them was rather limited. Of the three mammalian thyroids, the human thyroid was provided with less numerous somatostatin-containing cells than the others (Fig. 1). In all cases, somatostatin-containing cells were predominantly located in the parafollicular areas although scattered cells were also seen in the follicular areas (Fig. 3). Somatostatin-containing cells were oval, round or polygonal in shape, with strongly and homogeneously fluorescent cytoplasm and large non-fluorescent central nuclei. After incubation with somatostatin antibody absorbed with thyroxine, triiodothyronine or porcine calcitonin, immunofluorescent cells could still be seen. On the other hand, they

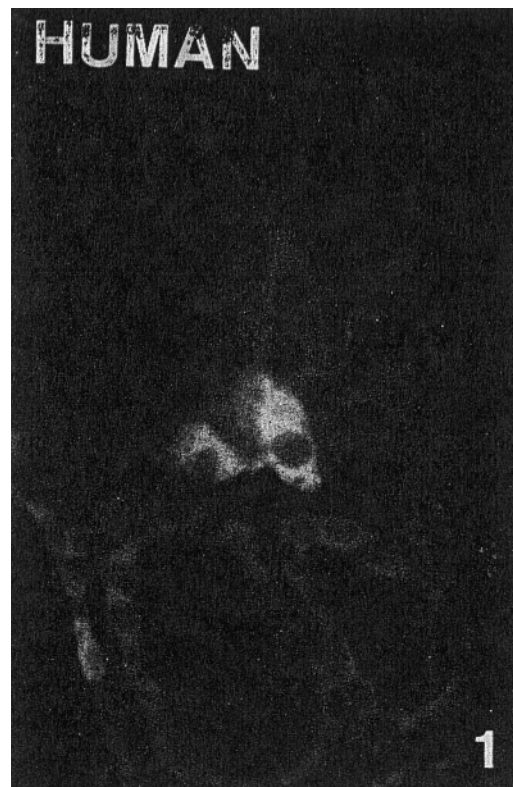
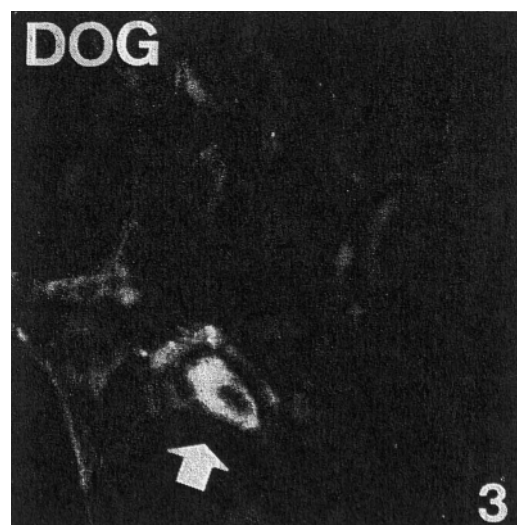


Fig. 1. Immunofluorescence micorgraph of the human thyroid after incubation with somatostatin antibody. Somatostatin-containing cells are seen in the interfollicular position.



Figs. 2 and 3. Immunofluorescence micrographs of the dog thyroid after incubation with somatostatin antibody. Somatostatin-containing cells are situated in the interfollicular (Fig. 2) and the follicular (Fig 3, arrow) positions.

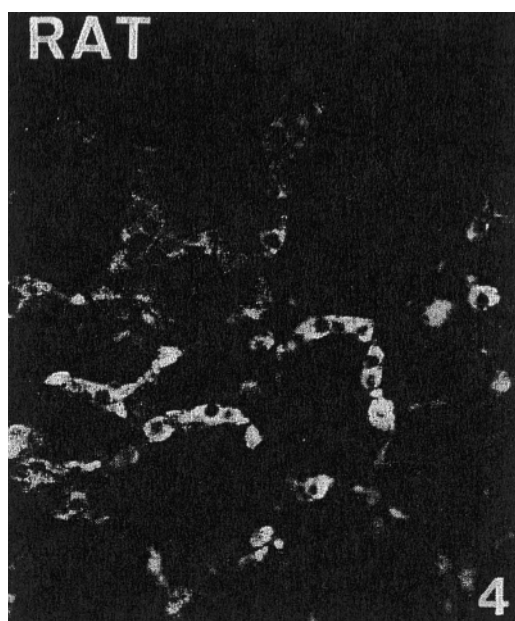


Fig. 4. Immunofluorescence micrograph of the rat thyroid after incubation with somatostatin antibody. Somatostatin-containing cells are seen in the interfollicular position.

were not detected after incubation with somatostatin antibody absorbed with synthetic somatostatin. These immunocytochemical reactions, therefore, were best explained by proposing that these cells contained somatostatin or somatostatin-like substances.

DISCUSSION

The present study demonstrates the occurrence of cells reacting with somatostatin antibody in the human, dog and rat thyroids. These results give immunohistochemical evidence for the presence of somatostatin, a hypothalamic tetradecapeptide, in the thyroid of mammals including human. These positive cells are distributed both in the interfollicular and in the follicular areas. This result agrees to an observation of Hökfelt et al. (1975) in the rat thyroid.

Now, it is known that in addition to the principal cells (epithelial or glandular cells) of the thyroid follicles, there is another small population of cells which are distributed in the follicular epithelium and interfollicular areas. The latter has commonly been called "parafollicular cells" and also referred to as "mitochondrion-rich cells" or "C cells" (Nonidez 1931/1932). They arise from cells in the epithelium and gradually withdraw into the interfollicular areas, so that many of the parafollicular cells are interfollicular in position (Bloom and Fawcett 1968). It was also established by immunofluorescence using the calcitonin antibody (Bussolati and Pearse 1967) that the parafollicular cells of the thyroid are the source of calcitonin. From the fact that the localization of somatostatin-containing cells in the thyroid well corresponds to that of parafollicular cells, it seems likely that the somatostatin-containing cells, as well as the calcitonin-containing cells, belong to the parafollicular cells.

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